

## **Prometaphase Chromosome Preparation from Mouse Spleen (C57Bl/6)**

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### **Reagents**

**Acetic acid, glacial**

**Antibiotic-Antimycotic 100x**

10,000 U/ml Penicillin G sodium, 10,000 µg/ml streptomycin sulfate, 25  
µg/ml amphotericin B

Invitrogen, Cat. 15240-013

**Bromodeoxyuridine (BrdU)**

Sigma, Cat. B9285

**Colcemid, KaryoMAX Colcemid Solution, 10 µg/ml**

Invitrogen, Cat. 15210-016

**Concanavalin A (5 µg/µl)**

Sigma, Cat. C-5275

**Fetal Bovine Serum (FBS) heat inactivated**

Invitrogen, 16140-022

**L-Glutamine-200 mM, 100x**

Invitrogen, 25030-016

**Homogenizer**

Thomas Scientific, Cat. 3431D7

**Lipopolysaccharides (LPS) 5mg**

Sigma, Cat. L-2637

**Methyl alcohol, anhydrous**

Mallinckrodt, Cat. 3016

**Methotrexate (MTX), 500 mg**

Sigma, Cat. M 8407

**Potassium chloride (KCl)**

**RPMI Medium 1640**

Invitrogen, Cat. 21870-050

## Preparation

### Reagents

	<b>Amount</b>
<b>Concanavalin A</b>	
Concanavalin A	5 mg
Sterile water	1 ml
For a stock solution of 5µg/µl	

### RPMI 1640 Complete Medium

<b>Components</b>	<b>Amount</b>
RPMI Medium 1640	440 ml
Antibiotic-Antimycotic, 100X	5 ml
L-Glutamine-200 mM, 100X	5 ml
Fetal Bovine Serum (FBS)	50 ml

### Fixative

Prepare fresh: methanol/acetic acid 3:1, volume:volume

### Hypotonic Solution: 0.075M KCl

KCl	5.6 g
Distilled water	1000 ml

### Lipopolysaccharides (LPS), stock solution

Lipopolysaccharides (LPS)	25 mg
Sterile water	1 ml

Use 1:1000 dilution for a final concentration of 25 µg/ml of culture

### MTX stock

Make an initial stock of 10<sup>-3</sup> M in H<sub>2</sub>O and then dilute to 10<sup>-5</sup> M  
Prepare fresh with each use.

### BrdU stock

1 mg/ml in distilled water  
Prepare fresh with each use.

## Procedure

1. Prepare tissue culture flasks. To one T75 flask, add:

Components	Amount
Prepared media	20 ml
Concanavalin A (5µg/µl)	30 µl
Lipopolysaccharides (LPS)	25 µl

2. Isolate spleen from mouse. Transport in sterile, unsupplemented RPMI 1640.

3. Place three spleens into a homogenizer with 3 ml of plain RPMI media. Grind well.
4. Transfer 0.5 ml of cell suspension to each T75 flask.
5. Incubate at 37°C for 24 hr. After 24 hr add 200 µl of MTX stock ( $10^{-5}$ M) to 20 ml of culture (MTX final concentration of  $10^{-7}$ M); mix well and incubate an additional 17 hr.
6. After 17 hr centrifuge the content of the flasks, remove the supernatant, and wash the pellet twice with plain media.
7. After the second wash resuspend the pellet in 20 ml of RPMI 1640 complete media and transfer to a T75 flask.
8. Add 500 µl of the BrdU stock (1mg/ml) to a final concentration of 25 µg/ml (minimize light exposure).
9. Incubate for 5 hr 30 min at 37°C.
10. For the last 10 min of the incubation add 20 µl of Colcemid stock (10 µg/ml) to a final concentration of 0.06 µg/ml .
11. Centrifuge cultures for 10 min.
12. Transfer to 50 ml centrifuge tubes and centrifuge at 1000 rpm for 10 min.
13. Remove supernatant.
14. Gently add 10 ml 0.075M KCl (prewarmed to 37°C) to each tube and resuspend pellet.
15. Incubate tubes at 37°C for 15 min.
16. Following incubation, add a few drops of freshly prepared fixative.
17. Centrifuge at 1200 rpm for 10 min.
18. Remove supernatant.
19. Wash pellet with freshly prepared fixative, at least 3 times.
20. Store pellet under fixative at -20°C until ready to prepare slides.